

X-ray Crystallographic Study of Covalently Modified Carboxypeptidase A by 2-Benzyl-3,4-epoxybutanoic Acid, a Pseudomechanism-Based Inactivator

Mikyung Yun, Chihyo Park, Sangsoo Kim,* Doohyun Nam, and Sung Chun Kim

R & D Center, Lucky Ltd., P.O. Box 10
Dae-Deog-Danji, Dae-jeon 305-343, Korea

Dong H. Kim*

Department of Chemistry and
Center for Biofunctional Molecules
Pohang Institute of Science and Technology
P.O. Box 125, Pohang 790-600, Korea

Received November 19, 1991

In recent years, zinc-containing metalloenzymes have received special attention because of the roles that they play in the etiology of many serious diseases;¹ thus they serve as target enzymes in the design of therapeutically useful inhibitors of enzymes.² In this respect, carboxypeptidase A (CPA), a well-studied Zn²⁺-containing metalloexopeptidase,⁴ bears an unusual importance because it serves as a model for many metalloenzymes.^{1,3,5}

Recently, we reported that 2-benzyl-3,4-epoxybutanoic acid (BEBA) is a novel type of CPA inhibitor which inactivates CPA irreversibly in high efficiency.⁶ This inactivator was shown to be a pseudomechanism-based inactivator as designed on the basis

of a proposed enzymic mechanism of CPA.⁶ At the time, however, its nature as an inactivator and the proposed mode of inhibition were solely based on the kinetic studies and the chemical rationale used for the design. Now this communication reports the single-crystal X-ray characterization of the inactivated CPA by BEBA at 2-Å resolution,⁷ showing clearly that the carboxylate of Glu-270 is indeed covalently modified by the inactivator.⁶ Furthermore, the stereochemistry of the effective BEBA is established as the 2*S*,3*R* configuration on the basis of the electron density in 2*F_o* - *F_c* maps.

The electron density map (Figure 1) obtained from a single crystal⁷ of inactivated CPA shows the presence of a continuous electron density between Glu-270 and the ring-cleaved BEBA to indicate clearly the formation of a covalent bond between the carboxylate of Glu-270 and BEBA at the 4-position with a concomitant ring opening. The length of the newly formed C-O bond was modeled to be 1.4 Å, a typical value for an ester linkage.¹⁰ The hydroxyl at the C₃ of the chemically transformed inhibitor is coordinated to Zn²⁺ with the bond length 2.1 Å and the C-O...Zn²⁺ angle 103°, replacing the water molecule that is coordinated to the Zn²⁺ in the native CPA. The position of the carboxylate has changed little upon this covalent bond formation. As expected, the aromatic ring of Tyr-248 moved to the "down" position as seen in structures of CPA-ligand complexes.¹¹ The phenyl ring of the inactivator is seated in the S₁' subsite, having close contacts with Leu-203 and Ile-243 in a perpendicular fashion on each side of the ring with distances of 3.9 and 3.5 Å, respectively. This may be the result of CH/π interactions. The existence and importance of such interactions have recently been pointed out by Nishio and Hirata.¹²

The stereochemistry⁷ of the inactivator deserves a special mention. We were surprised to learn that the stereochemistry of the C₂ stereogenic center is the *S* configuration because it corresponds to the *D* series, which is opposite to the known stereochemistry of CPA substrates.¹³ CPA is shown to have *L* stereospecificity.^{1,4} This reversal of stereochemistry became, however, apparent when molecular models of four possible stereoisomers of BEBA were examined in comparison with those of substrates. Thus, when the benzyl and the carboxylate moieties of each BEBA stereoisomer were superimposed onto the corresponding moieties of *N*-acetyl-*L*-phenylalanine, only the epoxy oxygen of the 2*S*,3*R* isomer, which is presumed to ligate to the Zn²⁺ at the active site, was positioned in close proximity to the amide oxygen of the substrate.

To our best knowledge, the present communication describes the first single-crystal X-ray structure of CPA covalently modified at the carboxylate of Glu-270, which has been implicated as the catalytic reaction site of CPA. Furthermore, it confirms the kinetic results reported in the previous communication⁶ and strongly supports the legitimacy of the rationale used in the design of BEBA as a CPA inactivator.⁶ Lastly, the present study bears a con-

(1) (a) *Zinc Enzymes*; Bertini, L., Luchinat, C., Maret, W., Zeppezauer, M., Eds.; Birkhauser Boston, Inc.: Boston, 1986. (b) *Zinc Enzymes*; Spiro, T. G., Ed.; Wiley: New York, 1983. (c) Hartsuck, J. A.; Lipscomb, W. N. In *The Enzymes Hydrolysis: Peptide Bonds*, 3rd ed.; Boyer, P. D., Ed.; Academic Press: New York, 1971; Vol. 3, Chapter 1.

(2) This aspect has been amply demonstrated by the rational design of captopril, an antihypertensive agent which lowers high blood pressure by inhibiting the catalytic activity of angiotensin-converting enzymes.

(3) (a) Ondetti, M. A.; Rubin, B.; Cushman, D. W. *Science* 1977, 196, 441-444. (b) Cushman, D. W.; Cheung, H. S.; Sabo, E. F.; Ondetti, M. A. *Biochemistry* 1977, 16, 5484-5491.

(4) (a) For a recent review, see: Christianson, D. W.; Lipscomb, W. N. *Acc. Chem. Res.* 1989, 22, 62-69 and references cited therein. (b) Christianson, D. W.; Lipscomb, W. N. In *Mechanistic Principles of Enzymic Activity*; Liebman, J. F., Greenberg, A., Eds.; VCH Publishers: New York, 1988; Chapter 1.

(5) (a) Kim, D. H.; Guinasso, C. J.; Buzby, G. C.; Herbst, D. R.; McCauly, R. J.; Wicks, T. C.; Wendt, R. L. *J. Med. Chem.* 1983, 26, 394-403. (b) Patchett, A. A.; Harris, E.; Tristram, E. W.; Wyratt, M. J.; Wu, M. T.; Taub, D.; Peterson, E. R.; Ikeler, T. J.; ten Broeke, J.; Payne, N. G.; Ondeyka, D. L.; Thorsett, E. D.; Greenlee, W. J.; Lohr, N. S.; Hoffsommer, R. D.; Joshua, H.; Ruyle, W. V.; Rothrock, J. W.; Aster, S. D.; Maycock, A. L.; Robinson, F. M.; Hirschmann, R. F.; Sweet, S. C.; Ulm, E. H.; Gross, D. M.; Vassil, T. C.; Stone, C. A. *Nature (London)* 1980, 288, 280-283. (c) Mock, W. L.; Tsay, J.-T. *Biochemistry* 1986, 25, 2920-2927. (d) Vallee, B. L.; Auld, D. S. *Proc. Natl. Acad. Sci. U.S.A.* 1990, 87, 220-224. (e) Gafford, J. T.; Skidgel, R. A.; Erdos, E. G.; Hersh, L. B. *Biochemistry* 1983, 22, 3265-3271.

(6) Kim, D. H.; Kim, K. B. *J. Am. Chem. Soc.* 1991, 113, 3200-3202.

(7) The BEBA complex was cocrystallized with chromatographically separated BEBA in a racemic form by dialysis as described for the native crystals.⁸ The BEBA-CPA crystals belong to space group *P2₁* with *a* = 65.5, *b* = 60.6, *c* = 74.4 Å, *β* = 98.2°, and two molecules in the asymmetric unit. Data were measured using a FAST system to 2-Å resolution from two crystals (*R*_{merge} = 0.071). The structure was solved using the molecular replacement method with the native structure⁹ as the starting model and was refined with X-PLOR.⁹ The stereochemistry and conformation of bound BEBA were determined unambiguously from well-defined electron density in 2*F_o* - *F_c* maps. The present model, including two CPAs, two BEBAs, and 384 water molecules, is characterized by an *R* factor of 0.18 for 29 062 reflections between 15 and 2 Å, with the deviations from ideality of 0.015 Å and 3.08° for bond lengths and bond angles, respectively. Crystallographic details shall be included in a forthcoming full paper.

(8) Rees, D. C.; Lewis, M.; Lipscomb, W. N. *J. Mol. Biol.* 1983, 168, 367-387.

(9) Brunger, A. T. *X-PLOR Manual, Version 2.1*, Yale University: New Haven, CT, 1991.

(10) Dunitz, J. D. *X-ray Analysis and Structure of Organic Molecules*; Cornell University Press: Ithaca, NY, 1979.

(11) (a) Reeke, G. N.; Hartsuck, J. A.; Ludig, M. L.; Quioco, F. A.; Steitz, T. A.; Lipscomb, W. N. *Proc. Natl. Acad. Sci. U.S.A.* 1967, 58, 2220-2226. (b) Rees, D. C.; Lipscomb, W. N. *J. Mol. Biol.* 1982, 160, 475-498.

(12) (a) Sakakibara, K.; Hirota, M. *Chem. Lett.* 1989, 921-924. (b) Nishio, M.; Hirota, M. *Tetrahedron* 1989, 45, 7201-7245.

(13) This kind of reversal of the stereochemistry of ligands has precedents in enzymic reactions.¹⁴

(14) (a) Hein, G. E.; McGriff, R. B.; Niemann, C. J. *Am. Chem. Soc.* 1960, 82, 1830-1831. (b) Cohen, S. G.; Crossley, J.; Khedouri, E.; Zand, R. *J. Am. Chem. Soc.* 1962, 84, 4163-4164. (c) Cohen, S. G.; Milovanovic, A.; Schultz, R. M.; Weinstein, S. Y. *J. Biol. Chem.* 1969, 244, 2664-2674. (d) Keinan, E.; Hafeli, E. K.; Seth, K. K.; Lamed, R. *J. Am. Chem. Soc.* 1986, 108, 162-169.

(15) (a) Makinen, M. W.; Kuo, L. C.; Dymowski, J. D.; Jaffer, S. *J. Biol. Chem.* 1979, 254, 356-366. (b) Makinen, M. W.; Fukuyama, J. M.; Kuo, L. C. *J. Am. Chem. Soc.* 1982, 104, 2667-2669. (c) Sander, M. E.; Witzel, H. *Biochem. Biophys. Res. Commun.* 1985, 132, 681-687.

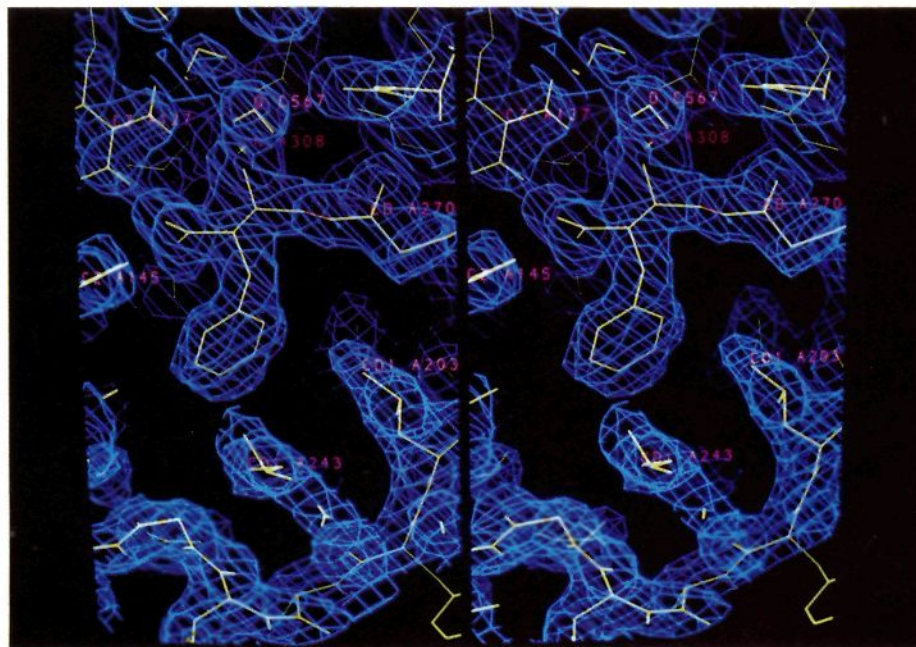


Figure 1. Stereoview of the electron density map and the final atomic model in the active site. The newly formed C–O bond between BEBA and Glu-270 of CPA is shown in red. The following residues are labeled: Arg-127, Arg-145, Leu-203, Ile-243, Glu-270, Zn^{2+} 308 (cross), and water 567.

siderable mechanistic significance,^{1,4} tending to support the anhydride mechanism of CPA.¹⁵ No covalent modification is expected to occur at Glu-270 by the alternative mechanism where the carboxylate of Glu-270 functions as a general base;^{1,4} instead, a diol may be produced.

Acknowledgment. We thank the Korea Institute of Science and Technology for the Cray time, and one of authors (D.H.K.) wishes to thank the Korea Science and Engineering Foundation for the partial support of this work and Kimoon Kim for helpful discussion.